Original article: Bacteriological profile of diabetic foot infections in a teritiary care teaching hospital

¹Y.Kavitha, ²S.Khaja Mohiddin

¹Assistant professor, Dept. of Microbiology , vinayaka mission medical college, karaikal, Puducherry , India ²Tutor, Dept. of Microbiology, vinayaka mission medical college, karaikal, Puducherry , India Corresponding author: S.khaja mohiddin Date of submission: 19 June 2014 ; Date of publication: 15 September 2014

Abstract:

Introduction: India has a diabetic population of about 50.8 million, which is expected to increase to 87 million by 2030.Diabetic foot infections are among the most common bacterial infections encountered in patients with diabetes mellitus. Hence, this study was carried out to determine the frequency of aerobic bacterial isolates from diabetic foot ulcers and their antibiogram.

Materials and methods: This was a prospective study conducted on clinical specimens which were taken from 56 patients with diabetic foot infections, over a six months period. The clinical specimens were processed by using the standard microbiological techniques. The anti-microbial susceptibility pattern was studied by the Kirby-Bauer disc diffusion method.

Results: Among 56 cases, 33(58.73%) had mono-microbial infections, 16(28.57%) had poly-microbial infections, and 7(12.5%) had sterile culture. Among bacteria isolated, 34(52.31%) were Gram negative and 31(47.69%) were Gram positive. All Gram negative bacilli showed good sensitivity to Imipenem,Piperacillin-Tazobactum and Amikacin. All Gram positive cocci remained 100% sensitive to Vancomycin followed by Amikacin and Clindamycin in a range of 71.4% to 100%.

Conclusion: Gram negative bacilli were predominantly isolated from diabetic foot ulcers. Piperacillin-Tazobactum and Amikacin would be essential for the empirical treatment.

Key words: Diabetic foot ulcers, Gram negative bacilli, Antibiogram

Introduction:

India has a diabetic population of about 50.8 million, which is expected to increase to 87 million by 2030.1 Among persons with diabetes mellitus, the risk of developing a foot ulcer is estimated to be 15%. Based on recent studies, the annual population based incidence ranges from 1.0% to 4.1% and the

prevalence range from 4% to 10%, suggesting the life time incidence as high as 25%.2

The impaired micro-vascular circulation in patients with a diabetic foot limits the access of phagocytes, thus favoring the development of an infection.3The local injuries and the improper footwear further compromise the blood supply in the lower extremi ties.4 While the foot infections in persons with diabetes are initially treated empirically, a therapy which is directed at the known causative organisms may improve the outcome.5 More than half of patients who have undergone lower extremity amputation will have a contralateral amputation within 5 years and half of those who undergo amputation will die within 3 years.6 E.coli, Klebsiella species, Proteus species, Pseudomonas species, S.aureus, and Enterococcus species are the most frequent pathogens which are cultured from diabetic foot ulcers.

The infections in the diabetic foot are usually polymicrobial due to aerobic bacteria, anaerobes and Candida spp. The severe infections usually yield polymicrobial isolates, whereas the milder infections are generally monomicrobial.7The specific organisms found in diabetic footinfections will differ not only from patient to patientand hospital to hospital, but also from one part of thecountry to another.Hence, this study was carried out to determine the frequency of aerobic bacterial isolates from diabeticfoot ulcers and their antibiogram.

Materials and Methods:

A prospective study was conducted on 56 patients admitted with infected diabetic foot over a period of six months(Jan 2010 to Jun 2010)at PES medical college and hospital, Kuppam. Proforma includes age, sex, history of trauma, habit of bare foot walking, alcohol, smoking,socio-economic status, duration of diabetes, duration of diabetic foot and location of foot ulcer.All ulcers were graded as per Wagner's classification.8 Diabetic foot ulcers of grade 1-5 were included and grade 0 and patient with limb amputation were excluded from the study.

Specimen collection: Wound beds were prepared before specimen collection, where the wound immediate surface exudates and contaminants were cleansed off with moistened sterile gauze and sterile normal saline solution. Dressed wounds were cleansed with non bacteriostatic sterile normal saline after removing the dressing. Aseptically the end of a sterile cotton-tipped applicator was rotated over 1 cm² area for 5 seconds with sufficient pressure to express fluid and bacteria to surface from within the wound tissue. Two swabs were collected from each patient, One swab was used for Gram staining and the other was used for culture. The specimens were inoculated on blood, chocolate and MacConkey agar plates and incubated aerobically for 24 to 48 hours at 37°c. Bacteriological culture and examination was done following standard microbiological techniques.9 Antibiotic sensitivity testing:

Antimicrobial susceptibility of the isolates was determined against the following antibacterial agents by Kirby Bauer disk diffusion method on Muller Hinton agar plates according to Clinical and Laboratory Standard Institute (CLSI) guidelines.¹⁰ Amikacin, Ceftazidime ,Cefotaxime, Ciprofloxacin, Cotrimoxazole, Gentamycin, Piperacillin-Tazobactum, Imipenem, Clindamycin, Erythromycin,Vancomycin,Cotrimaxazole, Amoxy-Clav, Ceftriaxone(Hi Media, Mumbai).

Results:

Among 56 patients with diabetic foot ulcers,41 were male and 15 were female and the age ranged from 35 -72 with mean age being 53 years. TABLE 1 Characteristics of diabetic foot specimens

Total Patients examined 56

Patients with microbial growth 49(87.5%)

No growth 7(12.5%)

Monomicrobial 33(58.73%)

Polymicrobial16(28.57%)

Gram Negative Isolates34(52.31%)

Gram Positive Isolates31(47.69%)

TABLE 2Bacteria isolated from diabetic foot infections

	Name of the isolate	Number of
S.NO		isolates(n=65)
1	Staphylococcus aureus	21(32.31%)
2	Klebsiellapneumonia	10(15.38%)
3	Pseudomonas aeruginosa	8 (12.31%)
4	Proteus mirabilis	6 (9.23%)
5	Proteus vulgaris	4(6.15%)
6	Enterococcus species	4(6.15%)
7	Coagulase negative staphylococci	4(6.15%)
8	Escherichia coli	3(4.62%)
9	Streptococcus pyogenes	2(3.08%)
10	Providencia species	2(3.08%)
11	Morganellamorganii	1(1.54%)

TABLE 3

Antimicrobial sensitivity pattern of Gram positive cocci.

ANTIBIOTIC	Stahylococcusaureus n=21	Enterococcus species n=4	Coagulase negative staphylococci n=4	Streptococcus pyogenes n=2
Gentamicin	8(38.1%)	1(25%)	2(50%	1(50%)
Amikacin	18(85.7%)	3(75%)	4(100%)	1(50%)
Vancomycin	21(100%)	4(100%)	4(100%)	2(100%)
Erythromycin	9(42.8%)	2(50%)	3(75%)	2(100%)
Clindamycin	15(71.4%)	3(75%)	4(100%)	2(100%)
Amoxy-Clav	12(57.1%)	2(50%)	3(75%)	2(100%)
Ciprofloxacin	13(61.9%)	0(0%)	2(50%)	1(50%)
Ceftriaxone	5(23.8%)	1(25%)	2(50%)	1(50%)
Cefotaxime	7(33.3%)	1(25%)	1(25%)	1(50%)

TABLE 4

Antimicrobial sensitivity pattern of Gram negative bacilli

Antibiotic	Klebsiella	Pseudomona	Proteus	Escherichi	Providenci	Morganellamorgani
	pneumoni	s aeruginosa	species	a coli	a species	i
	а	n=8	n=10	n=3	n=2	n=1
	n=10					
Gentamicin	5(50%)	3(37.5%)	6(60%)	1(33.3%)	1(50%)	1(100%)
Amikacin	7(70%)	5(62.5%)	8(80%)	3(100%)	2(100%)	1(100%)
Amoxy-Clav	7(70%)	6(75%)	9(90%)	3(100%)	2(100%)	1(100%)
Ciprofloxacin	6(60%)	3(37.5%)	5(50%)	2(66.6%)	2(100%)	1(100%)
Cotrimaxazole	4(40%)	3(37.5%)	4(40%)	1(33.3%)	2(100%)	1(100%)
Imipenem	10(100%)	6(75%)	10(100%	3(100%)	2(100%)	1(100%)
)			
Ceftriaxone	3(30%)	2(25%)	5(50%)	2(66.6%)	1(50%)	0(0%)
Piperacillin/Tazobactu	9(90%)	6(75%)	9(90%)	3(100%)	2(100%)	1(100%)
m						
Ceftazidime	4(40%)	3(37.5%)	4(40%)	1(33.3%)	2(100%)	0(0%)
Cefotaxime	3(30%)	3(37.5%)	5(50%)	2(66.6%)	1(50%)	1(100%)

Discussion:

Diabetic foot ulcer is one of the most common complication requiring hospitalization among diabetic patients. A diabetic foot infection is defined as any inframalleolar infection in a diabetic.

These include paronychia, cellulitis, myositis, abscesses, necrotizing fasciitis, septic arthritis, tendinitis, and osteomyelitis. The most common and classical lesion, however, is the infected diabetic "mal-perforans" foot ulcer.11Males were predominant in the study population 41(73.21%). This is in agreement with the study conducted by Gadepalli.B et.al 12In the present study the maximum number of patients with infected diabetic foot ulcers belonged to Wagner grade 3. Diabetic foot is known for poly-microbial infections.13, 14 But in study. monomicrobial infections our were predominated. This is in agreement with the study conducted by Dhanasekaranet al.15

In our study, Gram negative bacilli34(52.31%) were prevalent than more gram positive cocci31(47.69%).In previous reports, researchers have shown the predominance of Gram-positive infections.13However, if individual isolate is concerned, Staphylococcus aureus21(32.31%) was predominated, which was in accordance with the others findings.16, 17 Second most prevalent pathogen was Klebsiella pneumoniae10(15.38%) followed by Pseudomonas aeruginosa8 (12.31%).But other studies have demonstrated gram negative bacteria as the predominant pathogen.18, 19These discrepancies could be partly due to the differences in the causative organisms which occurred over time and the geographical variation or the types and the severity of the infections which were included in the studies.

Most of the Gram positive cocci were found to be highly resistant to cephalosporins, gentamicin, and erythromycin. But they showed good sensitivity to amikacin and clindamycin. All Gram positive cocci remained sensitive to Vancomycin. Most of the Gram negative bacilli were highly resistance to gentamicin, cotrimaxazole, ciproflaxacin and cephalosporins. All gram negative bacilli showed good response towards imipenem, amikacin and piperacillin/tazobactam. This is in agreement with the study conducted by Ozer B et al.19

The emergence of resistant strains represents a compounding problem standing against the efforts to prevent amputation as infection is the single most common cause of amputation. Even if the microorganism is sensitive to one particular antimicrobial, the drug is unlikely to attain therapeutic concentration at the site of infection because of virulence factors, such as hemolysins, proteases, and collagenases, as well as short-chain fatty acids, that cause inflammation, impede wound healing, and contribute to the chronicity of the infection.20, 21

Conclusion:

In our study, Gram negative bacilli predominantly caused diabetic foot infections.

But. if individual isolate is concerned. Staphylococcus aureus was predominated. Piperacillin- Tazobactum, and Amikacin would be essential for the empirical treatment. Patterns of microbial infections are not consistent in patients with diabetic foot infections and therefore repeated evaluation of microbial characteristics and their antibiotic sensitivity is necessary for selection of appropriate antibiotics.

References:

1. Shaw JE, Sicree RA, Zimmet PZ (2010) Global estimates of the prevalence of diabetes for 2010 and 2030.Diab Res ClinPract 87: 4-14.

2. SinghNalin, Armstrong David G, Lipsky Benjamin A. "TheJournal of American Medical Association", Edition 2005; 293, page No. 217-28

3. Sivaraman U, Kumar S, Joseph NM, Easow JM, Kandhakumari G. Microbiological study of diabetic foot infections. *Indian J Med Specialities*.2011; 2(1):12-17.

4. Pappu AK, Sinha A, Johnson A. Microbiological profile of diabetic foot ulcer. *Calicut Med Journal*.2011; 9(3):e:1-4.

5. Citron DM, Goldstein EJC, Merriam VC, Lipsky BA. Bacteriology of moderate to severe diabetic foot infections and invitro activity of antimicrobial agents.*J ClinMicrobiol*.2007; 45 (9):2819–28.

6. Smith SR, Reed JF. Prevalence of mixed infections in the diabetic pedal wound: a perspective based on a national audit. . Int J Low Extrem Wounds 2002; 1(2):125–128.

7.Raja NS. Microbiology of the diabetic foot infections in a teaching hospital in Malaysia: a retrospective study of 194 cases. *J MicrobiolImmunol Infect* 2007; 40(1): 39-44.

8. Wagner FW "The Diabetic Foot" Edition 1987, Volume 10, Page No. 163-72

9.ForbesBA,Sahm DF, Weissfeld AS. Overview of bacterial identification methods and strategies. Bailey and Scott's Diagnostic Microbiology,12th ed., chapter 13. St.Louis: Mosby; 2007: 216-47.

10. The Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility testing, twenty first informational supplement, M100-S21, *Clinical and Laboratory Standards Institute*, 2011.

11. Lipsky BA, Berendt AR, Deery HG, et al. Diagnosis and treatment ofdiabetic foot infections. Clin Infect Dis. 2004;39(7):885–910.

12. Gadepalli.B. Dhawan, V. Sreenivas, A. Kapil, A. C. Ammini, and R. Chaudhry 2006. A clinico-microbiological study of diabetic foot ulcers in an Indian tertiary care hospital. Diabetes Care 29:1727-32.

Abdulrazak A, Bitar ZI, Al-Shamali AA, Mobasher LA (2005) Bacteriological study of diabetic foot infections.
J Diabetes Complications 19: 138-141.

14. Anandi C, Aaguraja D, Natarajan V, Ramanatham M, Subramaniam CS, Thulasiram M, Sumithra S (2004) Bacteriology of diabetic foot lesions. Ind J Med Microbiol 22: 175-178.

15. Dhanasekaran G, Sastry G, Viswanathan M (2003). Microbial pattern of soft tissue infections in diabetic patients in South India. Asian J Diabet 5: 8-10.

16. Yoga R, Khairul A, Sunita K, Suresh C. Bacteriology of diabetic foot lesions. Med J Malaysia. 2006;61Suppl A:14-16.

17. Dang CN, Prasad YD, Boulton AJ, Jude ED. Methicillin resistantStaphylococcus aureus in the diabetic foot clinic: a worsening problem.Diabet.Med. 2003; 20:159–61.

18. Umadevi S, Kumar S, Joseph NM, EasowJ M, Kandhakumari G, Srirangaraj S, et al. Microbiological study of diabetic foot infections. *Indian Journal of Medical Specialities*2011; 2(1):12-17.

19. Ozer B, Kalachi A, Semerci E, Duran N, et al. Infections and aerobic bacterial pathogens in diabetic foot infections. *African J of Microbiol Research*;2010; 4(20): 2153-60.

20. Bowler PG, Davies BJ. The microbiology of infected and noninfected legulcers. Int. J. Dermatol. 1999; 38:573–78.

21. VonEiff C, Peters G, Heilmann C. Pathogenesis of infections due tocoagulase-negative staphylococci. Lancet Infect. Dis.2002; 2:677–85.